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Novel *IGF1R* Variants in Short Stature: Lessons from Two Patients and Outcome of Growth Hormone Therapy

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ABSTRACT

The growth hormone (GH)-insulin-like growth factor 1 (IGF1) axis is essential for the regulation of growth. IGF1 exerts its effects through the IGF1 receptor type 1 (IGF1R) that plays a pivotal role in fetal and postnatal growth. Pathogenic monoallelic *IGF1R* variants are known to cause pre- and postnatal growth restriction, often accompanied by normal or elevated serum IGF1 levels. Herein, the clinical and genetic characteristics of two cases with *IGF1R* novel variants, describing their growth patterns, endocrinological findings, and response to recombinant human GH (rhGH) therapy are presented. Case 1 was a 6.3-year-old boy, with birth weight of 2,500 g [-2.5 standard deviation score (SDS)] and a height of 101.5 cm (-3.2 SDS). Laboratory investigations revealed IGF1 and IGFBP3 levels of 117.8 ng/mL (0.9 SDS) and 4.55 µg/mL (1.3 SDS), respectively. Clinical exome sequencing (CES) identified a novel heterozygous c.3722+1G>A/p.(?) variant in the *IGF1R* (NM_000875.5) inherited from the mother. At 6.9 years of age, rhGH treatment was initiated at a dose of 0.035 mg/kg/day. The patient has been receiving rhGH for two years, achieving a height gain of +0.3 SDS per year, with an uneventful follow-up. Case 2 was a 3-year-old male with short stature and a history of being born small for gestational age (SGA) (-2.6 SDS). His height and weight were 70.0 cm (-2.1 SDS) and 8.8 kg (-1.1 SDS), respectively. He had a history of frequent respiratory infections. Pituitary hormone levels were normal, and he had no evidence of GH deficiency. CES revealed a novel heterozygous variant c.2275_2278 dup/p.(Ala760Glyfs*21) in the *IGF1R*. Identifying genetic causes of idiopathic short stature in SGA babies is important, as it facilitates more precise diagnoses, reduces unnecessary testing, and potentially enables targeted therapies. Our experience with rhGH therapy in one patient suggests a modest growth response, consistent with previous studies. However, elevated IGF1 levels during treatment highlight the importance of balancing therapeutic doses to optimize height gains without causing side effects.

Keywords: *IGF1R*, short stature, growth hormone, IGF1

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What is already known on this topic?

The growth hormone-insulin-like growth factor 1 (IGF1) axis is the major regulator of longitudinal growth. IGF1 and IGF2 act through the IGF receptor type 1 (IGF1R). Monoallelic *IGF1R* gene variants result in pre- and postnatal growth failure, developmental delay, and elevated serum IGF1 levels.

What this study adds?

Clinical and molecular genetic characteristics of two patients with short stature were described. Two novel IGF1R variants were reported, one splice-site and one nonsense.

Introduction

The growth hormone (GH)-insulin-like growth factor 1 (IGF1) axis plays a central role in human growth and metabolism (1). Among its components, the IGF1 receptor (IGF1R) mediates the effects of IGF1 and IGF2. These growth factors bind to IGF1R to promote growth both in fetal and postnatal life. Thus, dysfunctions in either the *IGF1* or *IGF1R* genes, may lead to pre- and postnatal growth restrictions (2). Structurally, IGF1R is akin to the insulin receptor and operates as a heterotetrameric transmembrane glycoprotein (3). Its alpha subunit ensures ligand binding, while the beta subunit facilitates intrinsic tyrosine kinase activity for signal transduction (4).

IGF1R (MIM*147370), located on chromosome 15q26.3, encompasses 21 exons. Pathogenic variants in this gene may occur in both monoallelic and biallelic forms, with the latter generally being associated with more severe clinical manifestations. Over 170 *IGF1R* variants have been documented, including missense/nonsense variants and gross deletions (5). Clinically, these variants are frequently associated with intrauterine growth restriction (IUGR), short stature, microcephaly, delayed bone age, developmental delay, and dysmorphic features, including a receding hairline, triangular face, long/smooth philtrum, thin upper lip, and full lower lip. However, the expression and severity of these traits can vary, and not all individuals present with all these features, reflecting the multifactorial nature of these characteristics. In adults with *IGF1R* defects, a thorough evaluation is essential, particularly concerning components of metabolic syndrome and hypogonadism (6).

Identifying *IGF1R* defects in short stature remains challenging because of the variability in patient selection and genetic methodologies. A comprehensive approach, integrating both single nucleotide variants (SNVs) and copy number variant (CNVs) analyses, is essential for accurate diagnosis (7). Rapid advances in high-throughput next-generation sequencing (NGS) techniques have facilitated the diagnosis of patients with short stature due to *IGF1R* defects. This has reclassified numerous cases previously defined as idiopathic short stature into distinct genetic disorders.

Herein, we present two patients with novel heterozygous *IGF1R* variants, detailing their clinical presentations and discussing the outcomes of recombinant human GH (rhGH) therapy in one patient.

Subjects and Methods

Clinical Evaluation

Height measurements were obtained using calibrated Harpenden stadiometers (Holtain, Crymch, UK), and weight was measured with standard equipment. Standard deviation scores (SDS) for all measurements were computed using growth charts specific to Turkish children (8). Small for gestational age (SGA) was defined as a birth weight (BW) or length more than two standard deviations below the mean for gestational age (9).

Hormonal Assays

Plasma GH levels were assessed using electrochemiluminescence immunoassay on a Roche Cobas platform (Roche Diagnostics İstanbul, Türkiye). IGF1 and IGF binding protein 3 (IGFBP3) were analyzed by immunoassays using the IMMULITE 2000 system (Siemens Healthcare İstanbul, Türkiye). IGF1 and IGFBP3 SDS were calculated using the online tool Child Metrics (10,11,12).

Molecular Analyses

After obtaining informed consent from the parents, DNA isolation was performed from peripheral blood samples under the standard protocols of the QIAamp DNA Mini kit (Qiagen, Hilden, Germany). Sequencing was performed using the Illumina NextSeq platform, and each patient was read at least 20X depth in the clinical exome sequencing (CES) panel. Bioinformatic analyses and variant calling were performed using the Sophia-DDM-V5.08 bioinformatics analysis program (SOPHIA Genetics, Rolle, Switzerland). This test evaluates the coding regions and exon-intron boundaries in the relevant genes. During the analysis, only pathogenic (P), likely pathogenic (LP), or variants of unknown significance were reported according to current scientific knowledge based on the ClinVar database (13). This database is constantly updated, and the data in the report is as of the date the report was written (2024); changes are possible

in the future. The interpretation of the variants was based on the American College of Medical Genetics and Genomics (ACMG) 2015 guideline (14). To assess the population frequency of the variants, data from gnomAD, the 1000 Genomes Project, dbSNP, and ExAC were utilized (15). CNVs were also examined with this analysis. Segregation analyses were performed for the parents.

Case Presentations

Clinical and molecular genetic characteristics of two patients from two unrelated families are described.

Patient 1 (P1)

A 6.3-year-old boy was referred for evaluation of short stature. He was born at term with a BW of 2,500 g (-2.5 SDS), which confirmed a diagnosis of being SGA. Parents were unrelated. His neurodevelopmental milestones were normal for age, except for delayed walking, which he achieved at three years.

At the time of assessment, his height was 101.5 cm (-3.4 SDS), and his weight was 15.1 kg (-2.8 SDS). Body proportions were normal for his age, and head circumference (HC) was -1.8 SDS. Midparental height (MPH) was -1.7 SDS, and his mother was notably short (148.0 cm, -2.6 SDS). Pubertal examination was Tanner stage I. He had distinctive facial features, including a long philtrum and retro-micrognathia. The bone age assessment was aligned with his chronological age,

and the skeletal survey was normal. The ophthalmological evaluation identified strabismus, for which a follow-up was recommended.

Laboratory results showed normal liver, renal, and thyroid function tests. His IGF1 and IGFBP3 levels were 117.8 ng/mL (0.95 SDS) and 4.55 µg/mL (1.3 SDS), respectively. GH stimulation test revealed a peak GH concentration of 14.4 ng/mL, ruling out GH deficiency. Chromosome analysis was 46,XY.

At 6.9 years of age, rhGH treatment was initiated at a dose of 0.035 mg/kg/day. The patient has been receiving rhGH for two years, achieving a height gain of +0.3 SDS per year, with an uneventful follow-up. Patient 1's mother was also evaluated for her short stature. The IGF1 and IGFBP3 levels were 251 ng/mL (-0.51 SDS) and 4.24 µg/mL, respectively. She had no evidence of insulin resistance, and her metabolic profile was normal. Clinical and laboratory findings of P1 at diagnosis and during rhGH treatment are summarized in Table 1 and Figure 1.

Patient 2 (P2)

A 3-year-old boy was referred for evaluation of growth failure. He was born at term with a BW of 2,300 g (-2.6 SDS). His perinatal history reported concerns about short femur length, though no definitive diagnosis was made at birth. His parents were unrelated.

Table 1. Clinical and laboratory findings of P1 at the time of diagnosis and during rhGH therapy

	At presentation	1 st year of rhGH	2 nd year of rhGH
Age, years	6.3	7.5	8.4
Height, SDS	-3.4	-3.1	-2.8
Height velocity, SDS	-	0.08	0.23
BMI, SDS	-0.62	-0.74	-0.9
Bone age, years	6.0	7.0	8.0
MPH SDS	-1.7		
Tanner stage	I	I	I
GH stimulation test (peak GH) (ng/mL)	14.4		
IGF1 (ng/mL) RR: 57.5-216.0	117.8	177.1	278.1
IGF1, SDS	0.95	2.42	5.0
IGFBP3 (µg/L) RR: 1.5-6.0	4.55	4.39	4.24
IGFBP3, SDS	1.29	2.42	0.74
Fasting glucose (mg/dL)	82.0	80.0	83.0
Fasting insulin (µIU/mL)	4.1	4.0	4.3
Triglyceride (mg/dL)	NA	NA	36.0
LDL Cholesterol (mg/dL)	NA	NA	63.0

GH: growth hormone; BMI: body mass index; SDS: standard deviation score; MPH: midparental height; IGF1: insulin like growth factor 1, IGFBP3: insulin like growth factor binding protein 3; NA: not available; RR: reference range; rhGH: recombinant human growth hormone; P1: patient 1

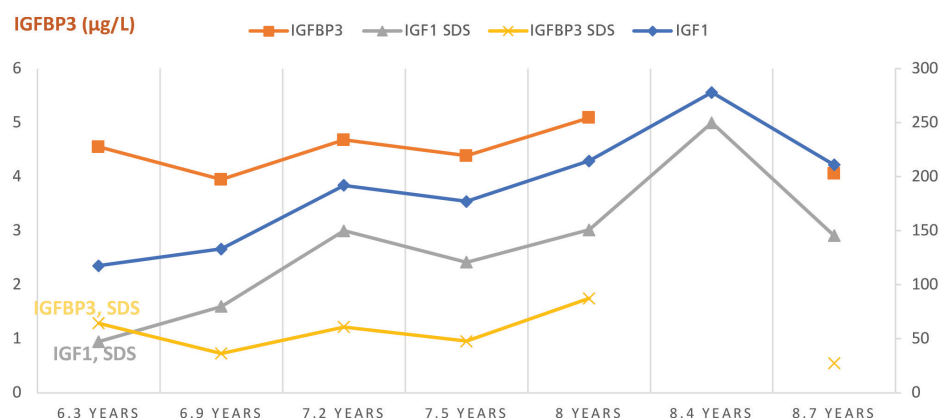


Figure 1. IGF1 and IGFBP3 levels of P1 while receiving GH treatment

IGF1: insulin like growth factor 1, IGFBP3: insulin like growth factor binding protein 3; P1: patient 1; GH: growth hormone; SDS: standard deviation score

At 1 year of age, his height was 70.0 cm (-2.1 SDS), weight was 8.8 kg (-1.1 SDS), and HC was 45.0 cm (-1.4 SDS). His mother was 158.0 cm (-0.87 SDS), his father was 166.0 cm (-1.65 SDS), and the MPH SDS was -1.25. The bone age corresponded to a chronological age of 3 to 6 months. Developmental milestones for gross motor and speech skills showed mild delays. Initial evaluations revealed normal IGF1 (88.53 ng/mL, -0.9 SDS) and IGFBP3 (4.2 ng/mL, -1.1 SDS). GH deficiency was ruled out with a glucagon stimulation test (peak GH: 8.0 ng/mL). The skeletal survey was normal.

At 2 years of age, his height velocity increased (+10.9 cm/year), however, height SDS remained below the expected range. At this time, his IGF1 level was elevated (164 ng/mL, 2.1 SDS), prompting further investigation for potential *IGF1R* variants.

His metabolic profile, including glucose (78 mg/dL), insulin (2.4 µIU/mL), triglyceride (35.5 mg/dL), and low density lipoprotein cholesterol (85 mg/dL), were normal.

The patient had recurrent respiratory infections, and selective immunoglobulin A deficiency was diagnosed, leading to a referral to clinical immunology for further evaluation.

At his most recent evaluation at 3.5 years old, his height and weight were 90.3 cm (-2.5 SDS) and 13.0 kg (-1.6 SDS), respectively. The arm span was 89.0 cm. The family was counseled about the possibility of initiating rhGH therapy.

The images of the patients and left-hand radiography are illustrated in Figure 2.

Molecular Genetic Results

Pedigree and Integrative Genomic Viewer of the NGS data are shown in Figure 3.

Analysis by CES in P1 revealed a heterozygous c.3722+1G>A/p.(?) variant in the *IGF1R* gene (NM_000875.5). This splice-site variant was classified as LP according to ACMG criteria (PVS1, PM2). This variant was classified as “deleterious” (MT, DANN, BayesDel) according to *in silico* prediction tools (16). For the splice-altering characteristics variant, SpliceAI is described as “Strong-Splice-altering” (17). This variant was not observed in the gnomAD (exomes and genomes) database (15). This *IGF1R* variant was also not reported in the HGMD professional database (November 2024) (5). Segregation analyses by Sanger sequencing revealed that his mother was also heterozygous for this variant.

In P2, the CES analysis identified a heterozygous c.2275_2278 dup/p.(Ala760Glyfs*21) variant in *IGF1R* (NM_000875.5), which is predicted to result in frameshift and premature termination. This variant was classified as LP according to ACMG criteria (PVS1, PM2) and was not observed in the gnomAD database (15). Once again, this *IGF1R* variant was also not reported in the HGMD professional database (November 2024) (5). Segregation analyses revealed that this variant was *de novo*.

Discussion

This report describes two rare cases of short stature due to novel heterozygous *IGF1R* variants, one involving a splice-site alteration and the other a frameshift variant. While the clinical features of these cases align with those previously reported, our findings contribute by documenting novel mutations and providing additional insights into growth response to rhGH therapy.

Although virtually all patients with monoallelic *IGF1R* variants present with pre- and postnatal growth restrictions, the extent varies remarkably. Walenkamp et al. (18) reported the detection of pathogenic *IGF1R* variants in approximately 2% of patients with short stature who were SGA. SGA-born cases represent a

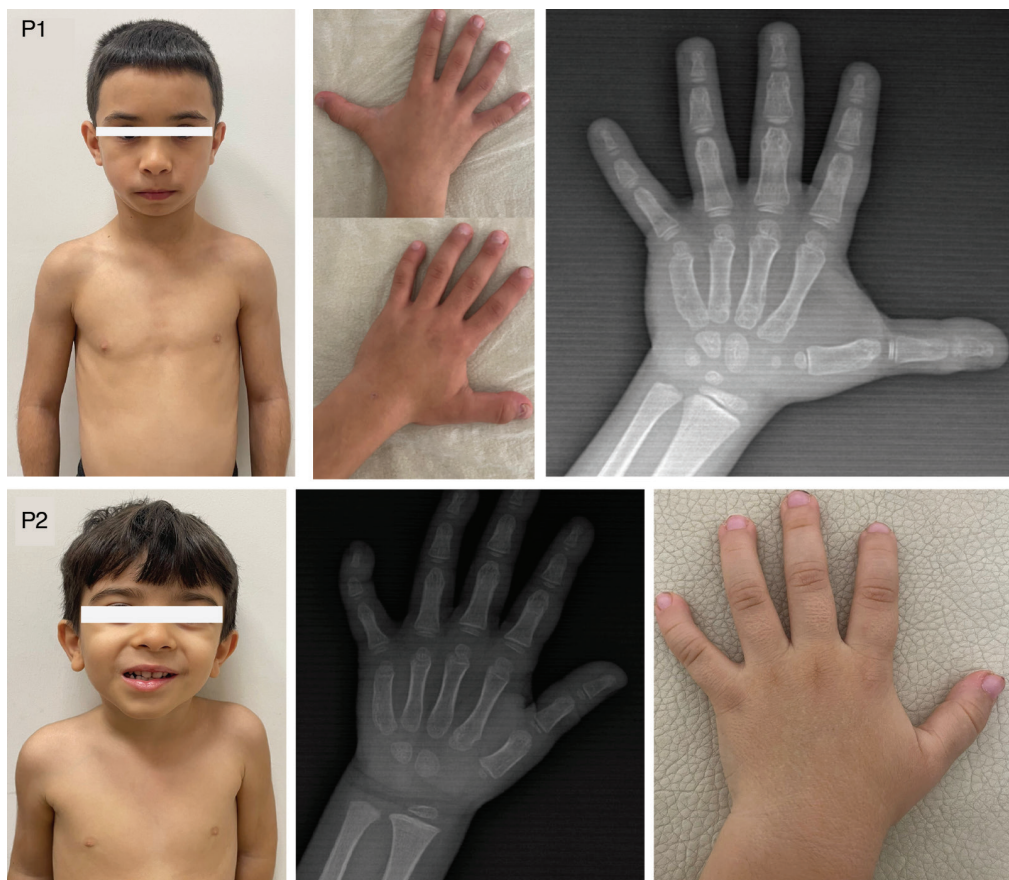


Figure 2. The pictures and left-hand radiography images of P1 and P2

P1: Long philtrum, retro-micrognathia, and clinodactyly. The bone age was consistent with chronological age without any obvious pathological appearance

P2: Long philtrum, micrognathia, mildly low-set ears, and clinodactyly. The bone age was consistent with chronological age with clinodactyly
P1: patient 1; P2: patient 2

diverse group with varying clinical features. The reduced size at birth can be attributed to fetal, maternal, placental, and/or genetic factors. While many SGA babies achieve normal growth by the age of 2 years, about 15% remain below -2.0 SDS in height and continue to be short. Genetic factors have been identified in a limited number of short SGA children, notably having point mutations and deletions in the *IGF1* and *IGF1R* genes (19). Klammt et al. (20) reported eight SGA patients who were in the range of -1.5 to -3.5 SDS due to *IGF1R* variants. Both of our patients were born SGA and were unable to catch up with growth by two years of age. P2 had a history of short extremities in the prenatal evaluation, but later postnatal examination revealed proportionate short stature.

In the study of Gonc et al. (7), patients with short stature without GH deficiency having either a low BW or microcephaly were evaluated to detect *IGF1R* defects, and variants were detected in 14% of the cohort. Two *IGF1R* deletions and five heterozygous

variants (one frameshift, four missense) were identified. All patients with *IGF1R* defects had a height, BW, and HC lower than -2.5 SDS, -1.4 SDS, and -1.36 SDS, respectively. IGF1 levels ranged from -2.44 to 2.13 SDS (7). Although the height SDS of P2 was similar, in our study, P1's height SDS at presentation was below -3.0 SDS.

Features described in cases with IGF1 resistance include mild facial dysmorphism (triangular face, brachycephaly, hypotelorism, low-set and prominent ears and micro-retrognathia), neurodevelopmental delay, and mild glucose intolerance (17). In our cases, some dysmorphic features, such as clinodactyly and micro-retrognathia, were also observed. Although the patients experienced delayed attainment of certain milestones, neuromotor development remained within normal limits. There were no abnormalities in carbohydrate metabolism. Given their prepubertal age, some clinical features may emerge later, and continuous monitoring will be necessary.

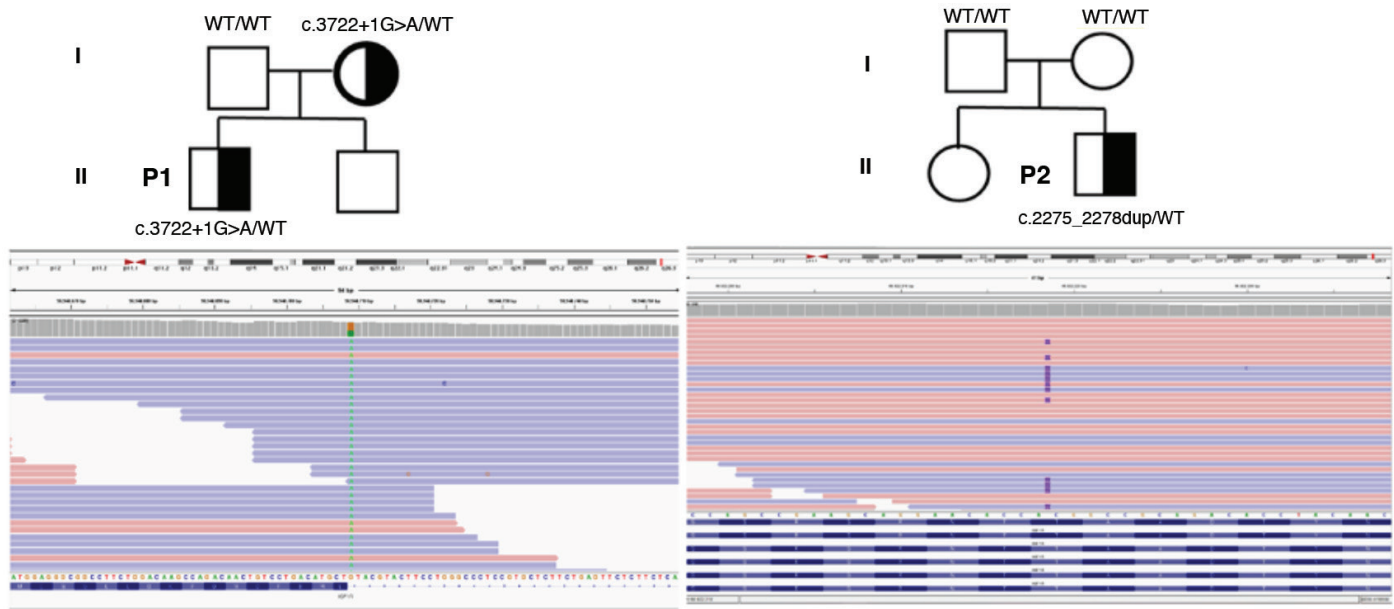


Figure 3. Pedigree and integrative genomics viewer (IGV) of the *IGF1R* gene variant in P1 and P2

The heterozygous (NM_000875.5) : c.3722+1G>A/p.(?) *IGF1R* variant in P1 and c. 2275_2278dup/p.(Ala760Glyfs*21) variant in P2. Segregation analysis was performed on the parents, while it was not conducted in clinically unaffected siblings. Future decisions regarding segregation analysis in siblings will be made based on their follow-up assessments

IGF1R: insulin-like growth factor 1 receptor; P1: patient 1; P2: patient 2

Cases of heterozygous *IGF1R* variants are typically characterized by IUGR, persistent postnatal growth failure, elevated serum IGF1 levels, and microcephaly (21). Our patients presented with moderately elevated IGF1 SDS values and did not exhibit microcephaly.

Most of the 170 variants identified in the *IGF1R* gene are missense/nonsense (n=107) and gross deletions (n=27). Splice-altering (n=9), small deletions (n=7), small indels (n=1), gross insertions (n=6), complex rearrangements (n=3), and regulatory region (n=3) variants have also been described (5). To date, splice-site variants (n=9) described are localized in the introns 1, 2, 3, 7, 8, 9, 10, and 18.

The c.3722+1G>A is a splice-site variant occurring at the donor site of intron 20. This variant is predicted to disrupt the normal splicing process, potentially leading to the exclusion of exon 20 from the mature mRNA transcript. Exon 20 encodes a portion of the tyrosine kinase domain of the IGF1R protein (catalytic domain of insulin receptor-like protein Tyrosine kinases), which is crucial for its signaling function. Alterations in this domain may impair the receptor's ability to transduce signals, affecting growth and development processes.

The *IGF1R* c.2275_2278dup/ p.(Ala760Glyfs*21) variant is a frameshift duplication located in exon 13 of the *IGF1R* gene (NM_000875.5). This exon encodes part of the extracellular

fibronectin type III domain, which is critical for ligand binding and proper receptor function (22). Variants in specific domains (e.g., fibronectin type III or tyrosine kinase) often lead to growth failure and altered IGF1 signaling, as seen in patients with short stature due to *IGF1R* variants (23).

Previous studies have highlighted significant variability in height, IGF1 levels, and intrauterine growth in individuals with the same heterozygous *IGF1R* variants. This variability is likely due to the complex structure of *IGF1R* and its interaction with the insulin receptor, which modulates IGF1 signaling. Although growth is influenced by multiple factors beyond IGF1 signaling, genetic testing for *IGF1R* alterations is crucial for accurate diagnosis, particularly in cases exhibiting elevated IGF1 levels or an exaggerated IGF1 response to rhGH therapy (24,25).

The phenotype of short stature associated with *IGF1R* variants is not fully elucidated, and no approved therapy is currently available. However, effectiveness of rhGH treatment has been studied (18). In this report, P1 was treated with rhGH, and the height SDS gain was +0.3 at the first year and +0.6 SDS at the second year. Patients with heterozygous *IGF1R* variants may respond to rhGH treatment, aligning with previous reports. However, the response remains variable (26). Çelik et al. (27) concluded in their recent review that rhGH has a partial beneficial effect in cases with *IGF1R* defects, particularly when initiated early and administered long-term. Nearly half of the

patients achieved a height gain of more than 1 SDS over the long term (26). Walenkamp et al. (18) suggested that higher IGF1 levels may need to be tolerated during the treatment to achieve a clinically significant increase in height SDS due to the partial insensitivity to IGF1 in these patients. The reluctance to increase the rhGH due to elevated IGF1 SDS levels may have contributed to suboptimal treatment response in our case. Higher IGF1 levels after rhGH therapy may indicate *IGF1R* gene variants (18).

The correlation between *IGF1R* defects and clinical presentation remains unclear, and the wide phenotypic variability complicates the selection of patients for *IGF1R* gene sequencing. However, advances in NGS technologies have enabled massively parallel sequencing of multiple genes and genomic regions with high precision, significantly enhancing diagnostic yield. By capturing both coding and non-coding regions, NGS facilitates the comprehensive identification of pathogenic variants, including SNVs, small in/dels, and CNVs, thereby revolutionizing the diagnostic approach to genetically heterogeneous disorders (28).

In conclusion, uncovering the genetic causes of idiopathic growth failure is important, as it facilitates more precise diagnoses, reduces unnecessary testing, and potentially enables targeted therapies. Our experience with rhGH therapy in one patient suggests a modest growth response, consistent with previous studies. A +0.6 SDS gain over two years aligns with prior reports of partial responsiveness in *IGF1R* variant carriers. However, elevated IGF1 levels during treatment highlight the importance of balancing therapeutic doses to optimize height gains without causing side effects.

Ethics

Informed Consent: Written informed consent was obtained from the patient's parents for the publication of details of their medical case and any accompanying images.

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Footnotes

Authorship Contributions

Surgical and Medical Practices: Mehmet Eltan, Hilal Sekizkardeş, Sezin Canbek, Murat Hakkı Yazar, Concept: Mehmet Eltan, Hilal Sekizkardeş, Sezin Canbek, Murat Hakkı Yazar, Saygın Abalı, Zehra Yavaş Abalı, Design: Mehmet Eltan, Zehra Yavaş Abalı, Data Collection or Processing: Mehmet Eltan, Hilal Sekizkardeş, Sezin Canbek, Murat Hakkı Yazar, Analysis or Interpretation: Mehmet Eltan, Sezin Canbek, Murat Hakkı Yazar, Literature Search: Mehmet Eltan, Saygın Abalı, Zehra Yavaş Abalı, Writing: Mehmet Eltan, Saygın Abalı, Zehra Yavaş Abalı.

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References

1. Miller BS, Rogol AD, Rosenfeld RG. The history of the insulin-like growth factor system. *Horm Res Paediatr*. 2022;95:619-630. Epub 2022 Nov 29
2. Klammt J, Pfäffle R, Werner H, Kiess W. IGF signaling defects as causes of growth failure and IUGR. *Trends Endocrinol Metab*. 2008;19:197-205.
3. De Meyts P. Insulin/receptor binding: the last piece of the puzzle? What recent progress on the structure of the insulin/receptor complex tells us (or not) about negative cooperativity and activation. *Bioessays*. 2015;37:389-397. Epub 2015 Jan 29
4. Adams TE, Epa VC, Garrett TP, Ward CW. Structure and function of the type 1 insulin-like growth factor receptor. *Cell Mol Life Sci*. 2000;57:1050-1093.
5. Stenson PD, Mort M, Ball EV, Shaw K, Phillips A, Cooper DN. The human gene mutation database: building a comprehensive mutation repository for clinical and molecular genetics, diagnostic testing and personalized genomic medicine. *Hum Genet*. 2014;133:1-9.
6. Abuzzahab MJ, Schneider A, Goddard A, Grigorescu F, Lautier C, Keller E, Kiess W, Klammt J, Kratzsch J, Osgood D, Pfäffle R, Raile K, Seidel B, Smith RJ, Chernašek SD; Intrauterine Growth Retardation (IUGR) Study Group. IGF-I receptor mutations resulting in intrauterine and postnatal growth retardation. *N Engl J Med*. 2003;349:2211-2222.
7. Gonc EN, Ozon ZA, Oguz S, Kabacam S, Taskiran EZ, Kiper POS, Utine GE, Alikasifoglu A, Kandemir N, Boduroglu OK, Alikasifoglu M. Genetic IGF1R defects: new cases expand the spectrum of clinical features. *J Endocrinol Invest*. 2020;43:1739-1748. Epub 2020 Apr 30
8. Neyzi O, Bundak R, Gökçay G, Günöz H, Furman A, Darendeliler F, Baş F. Reference values for weight, height, head circumference, and body mass index in Turkish children. *J Clin Res Pediatr Endocrinol*. 2015;7:280-293.
9. Kurtoğlu S, Hatipoğlu N, Mazıcıoğlu MM, Akın MA, Çoban D, Gökoğlu S, Baştuğ O. Body weight, length and head circumference at birth in a cohort of Turkish newborns. *J Clin Res Pediatr Endocrinol*. 2012;4:132-139. Epub 2012 May 4
10. Guven B, Can M, Mungan G, Acikgoz S. Reference values for serum levels of insulin-like growth factor 1 (IGF-1) and IGF-binding protein 3 (IGFBP-3) in the West Black Sea region of Turkey. *Scand J Clin Lab Invest*. 2013;73:135-140. Epub 2013 Jan 17
11. Demir K, Özen S, Konakçı E, Aydın M, Darendeliler F. A comprehensive online calculator for pediatric endocrinologists: ÇEDD Çözüm/TPEDS metrics. *J Clin Res Pediatr Endocrinol*. 2017;9:182-184. Epub 2017 Apr 26
12. Demir K, Konakçı E, Özkaya G, Kasap Demir B, Özen S, Aydın M, Darendeliler F. New features for child metrics: further growth references and blood pressure calculations. *J Clin Res Pediatr Endocrinol*. 2020;12:125-129.
13. Landrum MJ, Chitipiralla S, Kaur K, Brown G, Chen C, Hart J, Hoffman D, Jang W, Liu C, Maddipatla Z, Maiti R, Mitchell J, Rezaie T, Riley G, Song G, Yang J, Ziyabari L, Russette A, Kattman BL. ClinVar: updates to support classifications of both germline and somatic variants. *Nucleic Acids Research*. 2025;53:D1313-D21.
14. Richards S, Aziz N, Bale S, Bick D, Das S, Gastier-Foster J, Grody WW, Hegde M, Lyon E, Spector E, Voelkerding K, Rehml HL; ACMG Laboratory Quality Assurance Committee. Standards and guidelines for the interpretation of sequence variants: a joint consensus recommendation of the American College of Medical Genetics and Genomics and the Association for Molecular Pathology. *Genet Med*. 2015;17:405-424. Epub 2015 Mar 5
15. Chen S, Francioli LC, Goodrich JK, Collins RL, Kanai M, Wang Q, Alföldi J, Watts NA, Vittal C, Gauthier LD, Poterba T, Wilson MW, Tarasova Y, Phu W, Grant R, Yohannes MT, Koenig Z, Farjoun Y, Banks E, Donnelly S, Gabriel S, Gupta N, Ferriera S, Tolonen C, Novod S, Bergelson L, Roazen D, Ruano-Rubio V, Covarrubias M, Llanwarne C, Petrillo N, Wade G, Jeandet T, Munshi R, Tibbetts K; Genome Aggregation Database Consortium; O'Donnell-Luria A,

- Solomonson M, Seed C, Martin AR, Talkowski ME, Rehm HL, Daly MJ, Tiao G, Neale BM, MacArthur DG, Karczewski KJ. A genomic mutational constraint map using variation in 76,156 human genomes. *Nature*. 2024;625:92-100. Epub 2023 Dec 6
16. Schwarz JM, Cooper DN, Schuelke M, Seelow D. MutationTaster2: mutation prediction for the deep-sequencing age. *Nat Methods*. 2014;11:361-362.
17. Strauch Y, Lord J, Niranjana M, Baralle D. Cl-SpliceAI-Improving machine learning predictions of disease causing splicing variants using curated alternative splice sites. *PLoS One*. 2022;17:e0269159.
18. Walenkamp MJE, Robers JML, Wit JM, Zandwijken GRJ, van Duyvenvoorde HA, Oostdijk W, Hokken-Koelega ACS, Kant SG, Losekoot M. Phenotypic features and response to GH treatment of patients with a molecular defect of the receptor. *J Clin Endocrinol Metab*. 2019;104:3157-3171.
19. Ester WA, van Duyvenvoorde HA, de Wit CC, Broekman AJ, Ruivenkamp CA, Govaerts LC, Wit JM, Hokken-Koelega AC, Losekoot M. Two short children born small for gestational age with insulin-like growth factor 1 receptor haploinsufficiency illustrate the heterogeneity of its phenotype. *J Clin Endocrinol Metab*. 2009;94:4717-4727. Epub 2009 Oct 28
20. Klammt J, Kiess W, Pfäffle R. IGF1R mutations as cause of SGA. *Best Pract Res Clin Endocrinol Metab*. 2011;25:191-206.
21. Kawashima Y, Kanzaki S, Yang F, Kinoshita T, Hanaki K, Nagaishi J, Ohtsuka Y, Hisatome I, Ninomoya H, Nanba E, Fukushima T, Takahashi S. Mutation at cleavage site of insulin-like growth factor receptor in a short-stature child born with intrauterine growth retardation. *J Clin Endocrinol Metab*. 2005;90:4679-4687. Epub 2005 May 31
22. Turvey SJ, McPhillie MJ, Kearney MT, Muench SP, Simmons KJ, Fishwick CWG. Recent developments in the structural characterisation of the IR and IGF1R: implications for the design of IR-IGF1R hybrid receptor modulators. *RSC Med Chem*. 2022;13:360-374.
23. UniProt: the Universal Protein Knowledgebase in 2023. *Nucleic Acids Res*. 2023;51:D523-D531.
24. Juanes M, Guercio G, Marino R, Berenshtein E, Warman DM, Ciaccio M, Gil S, Bailez M, Rivarola MA, Belgorosky A. Three novel IGF1R mutations in microcephalic patients with prenatal and postnatal growth impairment. *Clin Endocrinol (Oxf)*. 2015;82:704-711. Epub 2014 Aug 7
25. Yang L, Xu DD, Sun CJ, Wu J, Wei HY, Liu Y, Zhang MY, Luo FH. IGF1R variants in patients with growth impairment: four novel variants and genotype-phenotype correlations. *J Clin Endocrinol Metab*. 2018;103:3939-3944.
26. Gkourogianni A, Andrade AC, Jonsson BA, Segerlund E, Werner-Sperker A, Horemuzova E, Dahlgren J, Burstedt M, Nilsson O. Pre- and postnatal growth failure with microcephaly due to two novel heterozygous IGF1R mutations and response to growth hormone treatment. *Acta Paediatr*. 2020;109:2067-2074. Epub 2020 Mar 6
27. Çelik NB, Losekoot M, Işık E, Gönç EN, Alikışifoğlu A, Kandemir N, Özön ZA. Long-term growth hormone therapy in a patient with *IGF1R* deletion accompanied by delayed puberty and central hypothyroidism. *J Clin Res Pediatr Endocrinol*. 2024;16:481-488. Epub 2023 Jan 23
28. Kruis T, Klammt J, Galli-Tsinopoulou A, Wallborn T, Schlicke M, Müller E, Kratzsch J, Körner A, Odeh R, Kiess W, Pfäffle R. Heterozygous mutation within a kinase-conserved motif of the insulin-like growth factor I receptor causes intrauterine and postnatal growth retardation. *J Clin Endocrinol Metab*. 2010;95:1137-1142. Epub 2010 Jan 26